

Journal of Advanced Zoology

ISSN: 0253-7214 Volume **44** Issue **5 Year 2023** Page **1063-1071**

Updates In Diagnosis And Management Of Genital Herpes

Amal Mustafa Ahmed Mohamed^{1*}, Abdulaziz Ali Salem Alqahtani², Asail Ahmed Alammar ³, Noora Abdulrahman Alrajhi ⁴, Noora Ahmed Juaythin⁵, Farah Fahad ALMuqrin⁶, Abdulmajeed Mohammed Alqahtani⁷, Hatim Musharraf Saeed Alamri⁸, Kholoud Abdullah AlHamdan⁹

^{1*}Obstetric and Gynecology Consultant, Abha Maternity and children hospital, MOH, Abha, Saudi Arabia
 ²General Practice Resident, Khamis Mushait Maternity and Children's Hospital (MOH), KSA.
 ³Medical Intern, College of medicine, Imam Mohammad Ibn Saud Islamic University, Riyadh, Saudi Arabia.

^{4,5,6}Medical Intern, College of medicine, Imam Mohammad Ibn Saud Islamic University, Riyadh, Saudi Arabia

^{7,8}Medical intern, College of medicine, King Khalid University, Abha, Saudi Arabia ⁹Medical Intern, College of medicine, Imam Mohammad Ibn Saud Islamic University, Riyadh, Saudi Arabia

*Corresponding Author: Amal Mustafa Ahmed Mohamed *Obstetric and Gynecology Consultant, Abha Maternity and children hospital, MOH, Abha, Saudi Arabia

Article History	Abstract
CC License	Herpes genitalia is mainly caused by herpes simplex virus type 2 and can appear as a primary or recurring infection. It is among the most prevalent sexually transmitted illnesses. This review article provides a comprehensive overview of the diagnosis, management, treatment, and prophylaxis of herpes genitalis, caused by herpes simplex virus type 2. It addresses critical areas of concern and aims to improve the often inadequate counseling and utilization of lab diagnoses, as well as provide updated information on treatment and management of the infection. This is a valuable resource for healthcare professionals and individuals seeking information on the pathogen and clinical manifestations of herpes genitalis.
CC-BY-NC-SA 4.0	Keyword: herpes genitalia, HSV-2, HSV -1, antiviral, vaccine, acyclovir

Introduction:

Genital herpes (Herpes genitalis) is a prevalent sexually transmitted disease its main cause is Herpes simplex virus (HSV) [1], type 1 and type 2 [2]. And it is marked by lifetime disease and occasional recurrence. An apparent breakout comprises solitary or cluster vesicles that ulcerate before settling on the the genitals perineum, buttocks, upper thighs, or perianal sites [3]. Because HSV is a chronic virus, there are significant concerns concerning the proper application of diagnostic methods, disease treatment, spread prevention to intimate partners, and suitable guidance [1]. HSV-2 causes Herpes genitalis in about 400 million people worldwide. In the United States of America according to The Centers for Disease Control and Prevention (CDC) [4]. In 2015-2016, 47.8% and 12.1% of US citizens aged 14-49 years were positive for HSV-1 and HSV-2, accordingly [4]. (HSV-2) is a highly common reason for herpes and one of the most frequent sexually transmitted diseases globally. (HSV-1) is primarily spread through asexual interaction throughout childhood. The majority of HSV-1 and HSV-2 viruses are asymptomatic (HSV-2) and affect about 22% of individuals aged 12 and above, amounting to 45 million people in the US alone [5]. While HSV-1 has been shown to induce genitalia infections in the perioral area, HSV-2 is more typically considered whenever individuals come *Available online at: https://jazindia.com*

with genitalia ulcers. Although this, the majority of breakouts of the illness will appear with vague signs such as vaginal itching, irritation, and excoriations, potentially delaying the start of therapy. Resulting in, more exposure to individuals with no infection possible most people don't recognize they are infected [6]. National Health and Nutrition Examination Survey study showed that only 13% of HSV-2 were positively diagnosed with herpes genitalia. When genital lesions are noticed type-specific molecular or virologic testing can be used to make an accurate diagnosis of genital herpes in addition to type-specific serology tests to detect when lesions aren't noticed [1]. Fatigue, a high fever, and localized adenopathy are all symptoms of primary infections. Later outbursts are typically weaker and are triggered by the resurrection of a dormant virus. Mostly, the traditional HSV appearance, whether primary infection or secondary outbreaks, is not present, with many patients experiencing minor or no symptoms. According to studies, 65% to 90% of people with genital HSV infection are ignorant of their infection [3]. After the first episode of infection, the viral DNA passes by axons to the spinal cord's sense ganglia, where it remains for the rest of the patient's life. When HSV is reactivated, it migrates back to the layer of skin and mucosa via an axon, its branches, or ipsilateral axons [7]. Unfortunately, The prevalence of genital herpes is increasing at an alarming rate [8]. It can cause clinical signs in the patient, but it can also cause asymptomatic viral infection [9], some antivirals have been used successfully for the management of symptomatic infections such as acyclovir which used as daily treatment up to 4 years as oppressive therapy [10]. Race, gender, and geography are all factors to consider. The number of lifetime sex partners is one of the risk factors as shown in Table 1 [3].

Risk factor	Risk increase/prevalence
Black, non-Hispanic	Three to four times the prevalence of non-Hispanic whites and Mexican-Americans
Female sex	Twice the prevalence of males
Hormonal contraception use, bacterial vaginosis, and vaginal group B streptococcus colonization	Each factor increases OR of shedding HSV-2: Hormonal contraception increases OR to 1.5 Bacterial vaginosis increases OR to 2.0 Group B streptococcus increases OR to 2.3
Number of lifetime sex partners	Increase in HSV-2 infection prevalence: 1 partner: 2% to 5% 2 to 4 partners: 7% to 19% 5 to 9 partners: 10% to 22% ≥ 10 partners: 19% to 37%
Oral-genital contact	Increases risk of transmitting HSV-1 from latent or active infection at oral site to genital site of uninfected partner
Presence of other sexually transmitted diseases	Increases OR of shedding to 3.1

Table 1 Risk factors for genital infections [3].

Diagnosis:

Herpes genitalia infection is primarily diagnosed clinically, particularly when the medical presentation is typical, with characteristic papular ulcers continuing to vesicles and ulcerated lesions that eventually crust, accompanied by local adenitis, and in relapsed cases preceded by prodromal. Yet, the clinical identification of genital herpes may be imprecise. The clinical distinction of genital HSV infection from other infections (Treponema pallidum, Haemphilusducreyi) and non-infectious reasons for genital lesions is frequently challenging, and laboratory verification of infection is generally recommended [11]. Herpes can be detected by antibodies in blood or by a viral infection in lesions. It can be detected by lab tests that include light microscopes, pharmacological analyses, virus culture, and serology & molecular methods or POC (point of

- care) Microfluidics-based assays that allow for immediate assessment [12]. HSV is also regularly diagnosed using human foreskin fibroblast (HEF), human heteroploid cell line (Hep-2), Medical Research Council cell strain 5 (MRC-5), primary rabbit renal tissue (PR), mink pulmonary, and green kidney cell strain (Vero Cells). After 2-3 days, the antigen from the virus should be detectable using quick immunoassay techniques such as direct immunofluorescence (IF) analysis or enzyme-linked immunosorbent assay (ELISA), followed by molecular biology techniques for HSV detection [13].
- **1- Light microscope:** HSV can be diagnosed with a simple microscopic test Mucocutaneous scrapping procedures (Tzanck smears) are obtained by unroofing vesicles and softly rubbing the vesicle bottom. These samples are treated with Giemsa, methylene blue, or Wright stain, and the appearance of cytopathic reactions (CPE) detects the HSV even though (Tzanck smears) are rapid and simple it is now infrequently as they have an inadequate sensibility and is unable to differentiate between HSV-1 and HSV-2 infection [14]. Used, Dacron swabs are collected on a glass slide treated with Methylene blue, Giemsa or Papanicolaou, or Hematoxylin and fixed in alcohol then examined by light microscope [13].
- **2- Electron microscopy:** Electron microscopy to examine vesicle fluid or other patient specimens for the determination of HSV has restrictions since virus appearance cannot be utilized to differentiate HSV from other herpes simplex viruses like a varicella-zoster virus. DFA dye, which can give type-specific distinction of HSV-1 and HSV-2, has virtually supplanted this old approach [15].
- **3- Direct Fluorescence Assay (DFA):** It's an appropriate substitute for the cell culture method as it dyes fixed cell samples with an accuracy of up to 90% in the early stages of infection [13].
- 4- Cell culture: The usual benchmark for HSV diagnosis and the standard method against which other methods are assessed is cell culture separation. While the test is 100% specific for HSV-1 or HSV-2, the accuracy is determined by the state of the infection at the time the sample is collected. The rate of response ranges as well, ranging from 75% for initial cases to 50% for relapse [15], spiecmen should be transported using a viral transport medium (VTM), which contains albumin and antimicrobials. Human foreskin fibroblasts and RK are the most commonly employed cell types because of their exceptional responsiveness in comparison to other cell types. CPE is discovered within 24 to 72 hours of primary injection, according to the responsiveness of the cell types, although the culture medium should be monitored for 7-10 days. The specificity of rabbit kidney and mink lung cell lines is greater (100% and 95%, respectively) than that of MRC-5 and Vero cell lines (77% and 64%, respectively). This procedure is tedious, produces limited sensibility, and necessitates specialized supplies, which is its principal shortcoming [13]. Herpes recuperation is the greatest impediment to high specificity in all culture-based and DFA assays. Herpes recuperation from ulcers may be quite diverse, depending on the current state of the disease and the initial vs recurring disease. The samples taken from vesicular-stage diseases have the greatest culture sensibility, with the virus isolated in as many as 95% of instances. When samples are acquired from ulceration (32 to 72%) and crusted (17 to 30%) infections, the test's specificity rapidly diminishes. Furthermore, lesions caused by the original infection might have a viral burden three to four orders of magnitude larger than reoccurring infections. Thus, viral recapture and consequent tolerance to culture from recurring HSV ulcers can be much lower than from original HSV ulcers [14].
- **5- Polymerase Chain Reaction:** It's the standard molecular diagnostic technique. The PCR approach, which uses a standard TaqMan probe or the HydProbe system to target particular areas of the genome, goes through numerous temperature cycling stages of amplification and produces results in the form of fluorescent signal information; an electrophoresis gel can also be employed. The benefits of fluorescence-based real-time PCR have radically transformed the PCR-based technology for DNA measurement [16]. It is extremely sensitive and precise, does not require any post-operation treatment, and may be controlled remotely. In immediate PCR, a fluorescent dye (SYBR green) or TaqMan probe is used to monitor DNA replication as the reaction continues. Various PCR tests have been utilized for identifying HSV over the years [17].
- **6- Serology method (Detection of Viral Antibodies in Blood):** The identification of antigens in circulation generated in response to the HSV is an alternate method of diagnosing HSV infection. This entails extracting serum from whole blood obtained from patients and applying one of the methods that follow to identify the existence of HSV-specific antigens [12].

- * Western blotting: (WB) assays for serologic identification and separation of HSV-1 and -2 are being established. Electrophoresis is used to extract whole-antigen compositions from HSV-1 or -2-diagnosed cells, which are then attached to a nitrocellulose gel and subjected to the patient's plasma. Applying bands of pattern particular to HSV-1 or -2 are used to assess HSV state [13-15].
- * Passive Agglutination or Hemagglutination Assay In the condition of a patient's serum specimen containing antibodies particular to the antigens, red blood cells bind with a water-soluble antigen on the outer layer and aggregates. HSV-1 glycoprotein C (gC-1) has been discovered as a significant virus a protein called with the capability to attach to rbcs during invasion. Antibodies towards this glycoprotein (gC-1 for HSV-1 and gC2 for HSV-2) have been found to emerge soon in the illness, making it an ideal candidate for identification of the virus [12,18]. The microtiter dish is loaded with rbcs that have been treated with gC-1 or gC-2 peptides. Within 3 hours, diluting sera from individuals with antibodies designed for gC-1 and gC-2 attach and aggregate RBCs. A diagnosis of infection with HSV is indicated by the creation of a distinct microscopic sequence of agglomeration. The microtiter dish is filled with rbcs, antigen (glycoproteins), and samples of serum in the hemagglutination suppression test. The antibodies in the specimen interact with the antigen, preventing red blood cells from agglutinating. The detection rate of the coagulation assay and the hemagglutination inhibition test was determined to be 97%, with 84% sensitivity, and the test requires around 2*10⁶ HSV particles/mL to get findings [12].
- * Early enzyme-linked immunosorbent assays (ELISAs): Whole-antigen formulations from HSV-1 or HSV-2- diseased cells were used. These tests are more precise (92 to 100%), less costly, and give a much faster response than WB. Nevertheless, a significant disadvantage is the low selectivity for HSV-1 and -2, which can vary from 61 to 85%. This is mostly owing to the inclusion of whole-antigen formulations in the construction of these tests. Because of the development of type-specific analyses that employ HSV glycoprotein G, raw antigen-based tests have become outdated [12]. SureVue-HSV-2 from Fisher (Houston, TX, USA), formerly known as the biokit HSV-2 fast diagnostic from Biokit [19], was the initial FDA-approved ELISA POC kit for HSV-2 identification. The technique uses capillary blood and serum to identify the HSV-2 gG2 genome. The assay's specificity ranged from 93.2% to 98.7%, and the accuracy was at 99.1%. Seroreversion of gG-specific antibodies can develop and can sometimes go undiagnosed, limiting the enduring validity of these diagnostic procedures. As a consequence, there is a risk of producing false-negative findings in these cases, having the virus potentially reactivating. Furthermore, the majority of these type-specific tests are incapable of detecting HSV-2 antibodies in HSV-1 seropositive patients [12,20].
- * Microfluidic-Based Point-of-Care Devices: The basic POC equipment consists of 1-a bio identification component enzymes, proteins, antibodies, and aptamer antibodies) that preferentially binds with the antigen and 2-a sensor that detects the contact and delivers either qualitative or quantitative results. The World Health Organization (WHO) has established particular rules for the development of point-of-care equipment for resource-limited programs which guarantee cost-effective, precise, sensitive to easy to use, quick, equipmentfree evaluation and transportation for on-site testing of samples to improve worldwide medical quality [12,21]. A capacity to offer quick assessment of specimens without the need of advanced equipment and with little or no professional oversight. Because of their affordable price, convenience, and ability to enable high-yield tests, microfluidic systems are being increasingly utilized for point-of-care equipment. Microfluidic systems often use tubes with dimensions smaller than 1000 microns, allowing for rapid distribution of reagents and specimens, turbulent flow, and huge surface-to-volume proportions, as well as allowing for testing with tiny specimen sizes [22]. Small tubes can be formed of any acceptable substance, including polymers, paper, glass, and fibers. The lateral flow immunoassay (LFIA) technique is a type of microfluidic innovation that is currently employed for the fast identification of HSV using nanoparticles such as golden nanoparticles/fluorescent nanoparticles linked to particular antibodies. Uni-GoldTM (Trinity Biotech Plc, Bray, Ireland) is the first publicly accessible FDA-approved microfluidic-based POC equipment for HSV-2 diagnosis [23]. The assessment has a response time of about 15 minutes and has been authorized for the widespread use and standard lab usage. If contrasted to the Western blot approach, the device's accuracy and precision are 94% and 99%, respectively [12].

Employing a dual-antigen immediate sandwich test, Aderman et al. established an equivalent type of lateral flow immunochromatographic assay (LFIA) employing gold nanoparticles linked with anti-HSV-2 IgG [24]. The created device's sensibility was determined to be 100% and its accuracy was determined to be 97.3%. This gadget has the potential to be used to construct a proof-of-concept assay for HSV-2 identification [12].

*Molecular-based assays: Several molecular methods are being employed to detect and measure viral DNA in various specimens. Amongst methods used in molecular biology, the nucleic acid amplification test (NAAT) is a highly precise tool for detecting HSV in intimate infection [25], asymptomatic infections of the genitals can also be detected with this technique. Real-time PCR and Enzyme Immunoassay, the combination are examples of molecular techniques. Compared to other procedures, Real time-PCR approach is widely employed because it gives precise data, is highly efficient, and has a small chance of contamination. PCR can identify viral genomes for some time after ulcers indicate genital herpes [15].

Management of Herpes Genitalia

Although many anti-herpesvirus drugs administering methods acyclovir, famciclovir, and valacyclovir are being investigated and are authorized by the FDA for initial medical incident genital herpes, periodic treatment, and inhibitory treatment, some therapies are less helpful to apply others because of their frequency of administration. Priority should be given to dosing techniques that are most likely to result in compliance by patients [1]. The majority of repeated HSV infections, whether oral or intimate, are minor and recover on their own. Resulting in the importance of therapy to decrease the length of the sickness, and antiviral drugs aid in this goal. Primary HSV disease, on the other hand, can cause generalized signs such as heat and exhaustion, along with lesional sores and lymphadenopathy. In these cases, antiviral medication is highly effective, but it can not avoid the reactivation of the persistent disease. Ongoing antiviral therapy has another benefit in individuals with herpes genitalia, namely avoiding relapse and decreasing the risk of transfer [26].

*Antiviral HSV-2 diseases are mostly treated with nucleoside analogs such as acyclovir (ACV), valacyclovir, and famciclovir. Most antivirals limit virus DNA polymerase action, stopping the virus from productively multiplying [27]. Dose of treatment of herpes genitalia is extremely adjustable, reliant on the development of the illness and the immunity of the patient [28]. Acyclovir is the first-line treatment for herpes genitalis. However, ingestion has an availability of just 15-30%. In normal immune response patients, herpes genitalis managed orally. Recurrent HSV infections ought to be administered intravenous (i. v.) acyclovir, especially in immunocompromised patients. Dose of acyclovir treatment for herpes genitalis is determined by the patient's viral situation, immunological competence, and whether or not she is pregnant. If relapse happens at a rate greater than 4-6 times per year prophylactic therapy is considered particularly during pregnancy [29-30]. Topical acyclovir is suggested for insignificant symptoms of herpes genitalis [31].

Valacyclovir is an acyclovir prodrug (an L-valyl ester) that can be taken orally. The liver enzyme valacyclovir hydrolase converts it to acyclovir after consumption. Orally it has a 54 percent bioavailability and achieves active component levels 3-4 times greater than oral acyclovir. This gives for longer dosing gaps and has been linked to improved adherence. Valacyclovir is also a conventional therapy for herpes genitalis in normal immune response patients, with studies demonstrating its efficiency in viral inhibition and preventing repeated herpes genitalis [28].

Famciclovir is the inactive diacetyl ester prodrug of penciclovir, the only topical efficient acyclic nucleotide analog formed by the splitting of 2 ester chains in the small intestine and hepatic. Orally has a bioavailability of 77%, one of the recommended therapeutic agents for herpes genitalis, however, it is not approved for use in young people, immunocompromised patients below 25, or pregnant women. So it should not be used as the primary treatment during pregnancy [32].

- *Essential Oils extracted from flowers in the Labiatae and Verbenaceae families have been found to have antiviral activity towards HSV. A study on the cells of Vero for HSV disease discovered that essential oils from the Labiatae had suppressive action with Rf values of 1*102 and 1*103 in the main illness [33]. Essential oil derived from Glechon spathulate and Glechon marifolia is highly efficient in genitalia herpes, using RC-37cells tea tree &eucalyptus oil has a virucidal effect, the lemon palm has antiviral effect. A study has shown the effectiveness of these oils on the HSV-2 virus [34].
- * Phytocompounds Several key flavonoids found in Houttuynia cordata, including quercetin, quercitrin, and isoquercetin, suppress HSV-2 more strongly [35]. Resveratrol is a plant flavonoid derived from Veratrum grandiflorum, inhibits the stimulation of NF-B. Resveratrol's antiviral efficacy against HSV-1 and HSV-2 has also been proven in experimental animals and cultivating cell experiments. Chikusetsusaponin Iva and calenduloside, both derived from the plant Alternanthera philoxeroides, were tested for antiviral efficacy towards HSV-1 and HSV-2 [36]. Glycopeptide meliacine (MA) from the plant Melia azedarach, when used topically HSV-2 antiviral efficacy in an animal sample [37].

*Vaccinations Four basic vaccine techniques are being established for preventing HSV-1 and HSV-2 diseases: replication-defective HSV vaccines, deactivated dead HSV vaccines, live-attenuated HSV vaccines, and subunit HSV vaccines. These vaccine techniques have benefits and drawbacks when it comes to effectiveness and security [36]. Vaccines classified as "therapeutic" or "prophylactic" vary in mechanism. Prophylactic vaccines are meant to avoid primary infection and consequent virus delay, while therapeutic vaccines seek to avoid recurring HSV illnesses and asymptomatic viral transmission in persons with latent HSV infection. HSV-2 became the primary focus of study in this area. Many vaccinations were investigated in the genital area animal models (mouse and guinea pig), but vaccines constructed from synthesized viral proteins appear to be the most likely to succeed at the moment. Adjunct HSV-2 protein component vaccinations and live attenuated HSV-2 deletion mutants were tested in randomized placebo-controlled studies [28]. There is optimism in the fact that primary infection defense is mostly provided by particular to the virus antibodies, whereas immunity generated by cells is more important in preventing relapse. Consequently, the immune system induced by the vaccine has to be greater than that induced by normal viral infection [38].

Inactivated killed vaccines work by using a virus that has been inactivated, or killed, so that it cannot cause disease. This virus is then used to stimulate the immune system to produce a protective immune response against the virus. In the case of HSV, the goal of an inactivated killed vaccine would be to stimulate the immune system to produce antibodies that can prevent infection with the virus. Several studies have been conducted to investigate the potential of inactivated killed HSV vaccines for genital herpes [4]. These studies have looked at the safety and efficacy of these vaccines in animal models and in early phase clinical trials in humans. While results have been promising in some cases, there are still challenges to overcome in the development of an effective inactivated killed HSV vaccine [38]. One challenge is the ability of the vaccine to stimulate a strong and long-lasting immune response. Inactivated killed vaccines may not be as effective at stimulating the immune system as live attenuated vaccines, which use a weakened form of the virus. Researchers are working to optimize the formulation and delivery of inactivated killed HSV vaccines to improve their ability to induce a protective immune response. Another challenge is the need for a vaccine that can provide protection against both HSV-1 and HSV-2, the two types of herpes simplex virus that can cause genital herpes. Developing a vaccine that can target both types of the virus is important for providing broad protection against genital herpes [17, 38]. Despite these challenges, inactivated killed HSV vaccines for genital herpes remain an area of active research. There is hope that continued research and development efforts will lead to the successful development of a safe and effective vaccine for preventing genital herpes. In the meantime, other prevention strategies such as antiviral medications and safe sexual practices continue to be important for reducing the spread of HSV [38].

Live-attenuated vaccines are a type of vaccine that contains a weakened form of the virus. When administered, the vaccine stimulates the immune system to produce a protective response against the virus, without causing the disease itself. This type of vaccine has been successful in preventing other viral infections, such as measles, mumps, and rubella [39]. Research into live-attenuated vaccines for genital herpes has shown promising results in animal studies. These vaccines have been shown to be effective in preventing infection and reducing the severity of symptoms in animals infected with HSV. Additionally, studies have demonstrated that these vaccines can stimulate a strong immune response, leading to the production of antibodies that can neutralize the virus [40]. However, there are still challenges and concerns surrounding the development of live-attenuated vaccines for genital herpes. One major concern is the potential for the weakened virus in the vaccine to revert to a more virulent form and cause disease in vaccinated individuals. Additionally, there is a risk of the vaccine causing infection in individuals with weakened immune systems. Despite these challenges, researchers continue to explore the potential of live-attenuated vaccines for genital herpes. Clinical trials are ongoing to assess the safety and efficacy of these vaccines in humans. If successful, these vaccines could provide a new and effective tool for preventing genital herpes and reducing the burden of this common and often stigmatized infection [39, 40].

Replication-defective virus vaccines work by using a modified form of the virus that is unable to replicate in the body. This means that the vaccine can stimulate an immune response without causing the disease. The goal of these vaccines is to train the immune system to recognize and attack the virus, thereby preventing or reducing the severity of genital herpes infections [41]. Several research studies have been conducted to evaluate the effectiveness of replication-defective virus vaccines for genital herpes. These studies have shown promising results, with some vaccines demonstrating the ability to stimulate a strong immune response and reduce the frequency of outbreaks in clinical trials. However, more research is needed to fully understand the long-term effectiveness and safety of these vaccines [42]. One of the challenges in developing replication-defective virus vaccines for genital herpes is the need to balance safety and efficacy. Since these vaccines use

a modified form of the virus, there is a risk of potential side effects or unintended consequences. Researchers are working to address these concerns by conducting rigorous preclinical and clinical studies to ensure the safety and effectiveness of these vaccines. In addition to safety and efficacy, another important consideration in the development of replication-defective virus vaccines for genital herpes is the need for widespread access and affordability. It is crucial that these vaccines are accessible to all individuals who are at risk of genital herpes, regardless of their socioeconomic status or geographic location. This will require collaboration between researchers, public health organizations, and policymakers to ensure that these vaccines are affordable and available to those who need them [41, 42].

Subunit HSV vaccines work by stimulating the immune system to produce antibodies that can recognize and neutralize the virus. By targeting specific parts of the virus, these vaccines can potentially provide more targeted and effective protection against HSV. Additionally, subunit vaccines may be better suited for individuals with weakened immune systems, who may not be able to receive live or attenuated virus vaccines [43]. Several subunit HSV vaccines are currently in development, and early clinical trials have shown promising results. These vaccines have been shown to be safe and well-tolerated, and have demonstrated the ability to stimulate an immune response against HSV. While more research is needed to determine their effectiveness in preventing genital herpes, the early results are encouraging [43]. It's important to note that the development of vaccines takes time, and it may be several years before subunit HSV vaccines are available for widespread use. In the meantime, it's important for individuals to continue practicing safe sex and to seek medical treatment if they suspect they have been exposed to HSV. Additionally, individuals with genital herpes should work closely with their healthcare providers to manage their symptoms and reduce the risk of transmitting the virus to others [19].

Nucleic acid vaccines are a type of vaccine that work by introducing genetic material from the virus into the body, which then triggers an immune response. This type of vaccine has shown promise in preclinical studies for genital herpes, as it has the potential to stimulate a strong and long-lasting immune response against the virus [42]. One of the advantages of nucleic acid vaccines is that they can be designed and produced relatively quickly, which makes them a promising option for responding to outbreaks of infectious diseases. Additionally, they are considered to be safe, as they do not contain live virus and cannot cause the disease they are designed to protect against [44]. Several nucleic acid vaccines for genital herpes have been developed and are currently being studied in clinical trials. These trials aim to evaluate the safety and effectiveness of the vaccines in preventing genital herpes outbreaks and reducing viral shedding. Early results from these trials have shown promising immune responses in participants, which suggests that nucleic acid vaccines could be a viable option for preventing and managing genital herpes in the future [45]. It is important to note that while nucleic acid vaccines for genital herpes show promise, they are still in the early stages of development and more research is needed to determine their long-term safety and effectiveness. Additionally, it will be important to address any potential challenges related to vaccine distribution, cost, and accessibility [44].

Conclusion:

Herpes is an STD illness that affects the lips and the genitalia. Nevertheless, some individuals are misdiagnosed, assuming they're suffering from HSV while they are healthy, and receive treatment with unnecessary over-the-counter medications. Many patients assume they are suffering from a "chronic" infection; nevertheless, HSV is a worldwide infection affecting nearly half of the world's populace, and genital herpes-suppressing medication is available. To aid patients, accurate diagnosis, thorough explanation, and dispelling myths are as vital as antiviral medication.

References:

- 1. Johnston C. (2022). Diagnosis and Management of Genital Herpes: Key Questions and Review of the Evidence for the 2021 Centers for Disease Control and Prevention Sexually Transmitted Infections Treatment Guidelines. *Clinical infectious diseases: an official publication of the Infectious Diseases Society of America*, 74(Suppl 2), S134–S143. https://doi.org/10.1093/cid/ciab1056
- 2. Mathew, R., Najeem, B., Sobhanakumary, K., Sunny, B., Pinheiro, C., & Anukumar, B. (2018). Herpes Simplex Virus 1 and 2 in Herpes Genitalis: A Polymerase Chain Reaction-Based Study from Kerala. *Indian journal of dermatology*, 63(6), 475–478. https://doi.org/10.4103/ijd.IJD_187_17
- 3. Groves M. J. (2016). Genital Herpes: A Review. American Family Physician, 93(11), 928–934.
- 4. Cole S. (2020). Herpes Simplex Virus: Epidemiology, Diagnosis, and Treatment. *The Nursing Clinics of North America*, 55(3), 337–345. https://doi.org/10.1016/j.cnur.2020.05.004

- 5. Xu, F., Sternberg, M. R., Kottiri, B. J., McQuillan, G. M., Lee, F. K., Nahmias, A. J., ... & Markowitz, L. E. (2006). Trends in herpes simplex virus type 1 and type 2 seroprevalence in the United States. *Jama*, 296(8), 964-973.
- 6. Mathew Jr, J., & Sapra, A. (2022). Herpes simplex type 2. *StatPearls [Internet]*.
- 7. Tata, S., Johnston, C., Huang, M. L., Selke, S., Magaret, A., Corey, L., & Wald, A. (2010). Overlapping reactivations of herpes simplex virus type 2 in the genital and perianal mucosa. *The Journal of Infectious Diseases*, 201(4), 499-504.
- 8. Leung, D. T., & Sacks, S. L. (2000). Current recommendations for the treatment of genital herpes. *Drugs*, 60, 1329-1352.
- 9. Martín, J. M., Villalón, G., & Jordá, E. (2009). Update on the treatment of genital herpes. *Actas Dermo-Sifiliográficas (English Edition)*, 100(1), 22-32.
- 10. Wong, S. S., & Yuen, K. Y. (2008). Antiviral therapy for respiratory tract infections. *Respirology (Carlton, Vic.)*, *13*(7), 950–971. https://doi.org/10.1111/j.1440-1843.2008.01404.x
- 11.LeGoff, J., Péré, H., & Bélec, L. (2014). Diagnosis of genital herpes simplex virus infection in the clinical laboratory. *Virology journal*, 11(1), 1-17.
- 12. Nath, P., Kabir, M. A., Doust, S. K., & Ray, A. (2021). Diagnosis of herpes simplex virus: laboratory and point-of-care techniques. *Infectious Disease Reports*, 13(2), 518-539.
- 13. Mazji, H. S., Meshkat, Z., Rezayi, M., Rezaee, S. A., Aryan, E., Gouklani, H., ... & Jalili, A. What is the best laboratory method for diagnosis of Herpes Simplex Virus in genital infections? *Archives of medical laboratory sciences*, 4(3).
- 14. Anderson, N. W., Buchan, B. W., & Ledeboer, N. A. (2014). Light microscopy, culture, molecular, and serologic methods for detection of herpes simplex virus. *Journal of Clinical Microbiology*, 52(1), 2-8.
- 15. Singh, A., Preiksaitis, J., & Romanowski, B. (2005). The laboratory diagnosis of herpes simplex virus infections. *Canadian Journal of Infectious Diseases and Medical Microbiology*, *16*, 92-98.
- 16.Deepak, S. A., Kottapalli, K. R., Rakwal, R., Oros, G., Rangappa, K. S., Iwahashi, H., ... & Agrawal, G. K. (2007). Real-time PCR: revolutionizing detection and expression analysis of genes. *Current genomics*, 8(4), 234-251.
- 17. Schremser, V., Antoniewicz, L., Tschachler, E., & Geusau, A. (2020). Polymerase chain reaction for the diagnosis of herpesvirus infections in dermatology: analysis of clinical data. *Wiener klinische Wochenschrift*, 132, 35-41.
- 18. Samae, M., Chatpun, S., & Chirasatitsin, S. (2021). Hemagglutination Detection with Paper-Plastic Hybrid Passive Microfluidic Chip. *Micromachines*, *12*(12), 1533. https://doi.org/10.3390/mi12121533
- 19. Morrow, R. A., Friedrich, D., Meier, A., & Corey, L. (2005). Use of biokitHSV-2 Rapid Assay to improve the positive predictive value of Focus HerpeSelect HSV-2 ELISA. *BMC Infectious Diseases*, 5(1), 1-7.
- 20. Ashley, R. L. (2001). Sorting out the new HSV type-specific antibody tests. *Sexually transmitted infections*, 77(4), 232-237.
- 21. Nath, P., Kabir, A., Khoubafarin Doust, S., Kreais, Z. J., & Ray, A. (2020). Detection of bacterial and viral pathogens using photonic point-of-care devices. *Diagnostics*, *10*(10), 841.
- 22. Sachdeva, S., Davis, R. W., & Saha, A. K. (2021). Microfluidic point-of-care testing: commercial landscape and future directions. *Frontiers in Bioengineering and Biotechnology*, *8*, 602659.
- 23. Shevlin, E., & Morrow, R. A. (2014). Comparative performance of the Uni-Gold[™] HSV-2 Rapid: a point-of-care HSV-2 diagnostic test in unselected sera from a reference laboratory. *Journal of Clinical Virology*, 61(3), 378-381.
- 24.Laderman, E. I., Whitworth, E., Dumaual, E., Jones, M., Hudak, A., Hogrefe, W., ... & Groen, J. (2008). Rapid, sensitive, and specific lateral-flow immunochromatographic point-of-care device for detection of herpes simplex virus type 2-specific immunoglobulin G antibodies in serum and whole blood. *Clinical and Vaccine Immunology*, 15(1), 159-163.
- 25.LeGoff, J., Péré, H., & Bélec, L. (2014). Diagnosis of genital herpes simplex virus infection in the clinical laboratory. *Virology journal*, 11(1), 1-17.
- 26.Imafuku, S. (2023). Recent advances in the management of herpes simplex in Japan. *The Journal of Dermatology*, 50(3), 299-304.
- 27. Majewska, A., & Mlynarczyk-Bonikowska, B. (2022). 40 Years after the Registration of Acyclovir: Do We Need New Anti-Herpetic Drugs?. *International journal of molecular sciences*, 23(7), 3431. https://doi.org/10.3390/ijms23073431
- 28. Sauerbrei, A. (2016). Herpes genitalis: diagnosis, treatment and prevention. *Geburtshilfe und Frauenheilkunde*, 76(12), 1310-1317.
- 29. German, S. T. I. (2016). Society. STI-treatment pocket guide.

- 30.Hollier, L. M., & Wendel, G. D. (2008). Third-trimester antiviral prophylaxis for preventing maternal genital herpes simplex virus (HSV) recurrences and neonatal infection. *Cochrane Database of Systematic Reviews*,).
- 31. Gross, G. (2004). Herpes simplex virus infections. *Der Hautarzt*, 55, 818-830.
- 32.Kang, S. H., Chua-Gocheco, A., Bozzo, P., & Einarson, A. (2011). Safety of antiviral medication for the treatment of herpes during pregnancy. *Canadian Family Physician*, *57*(4), 427-428.
- 33.Brand, Y. M., Roa-Linares, V. C., Betancur-Galvis, L. A., Durán-García, D. C., & Stashenko, E. (2016). Antiviral activity of Colombian Labiatae and Verbenaceae family essential oils and monoterpenes on Human Herpes viruses. *Journal of Essential Oil Research*, 28(2), 130-137.
- 34. Allahverdiyev, A., Duran, N., Ozguven, M. E. N. Ş. U. R. E., & Koltas, S. (2004). Antiviral activity of the volatile oils of Melissa officinalis L. against Herpes simplex virus type-2. *Phytomedicine*, 11(7-8), 657-661.
- 35.Chen, X., Wang, Z., Yang, Z., Wang, J., Xu, Y., Tan, R. X., & Li, E. (2011). Houttuynia cordata blocks HSV infection through inhibition of NF-κB activation. *Antiviral Research*, 92(2), 341-345.
- 36.Egan, K., Hook, L. M., LaTourette, P., Desmond, A., Awasthi, S., & Friedman, H. M. (2020). Vaccines to prevent genital herpes. *Translational Research*, 220, 138-152.
- 37. Petrera, E., & Coto, C. E. (2009). The therapeutic effect of meliacine, an antiviral derived from Melia azedarach L., in mice genital herpetic infection. *Phytotherapy Research*, 23(12), 1771-1777.
- 38. Halford, W. P., Geltz, J., Messer, R. J., & Hasenkrug, K. J. (2015). Antibodies are required for complete vaccine-induced protection against herpes simplex virus 2. *PLoS One*, *10*(12), e0145228.
- 39.Stanfield BA, Kousoulas KG, Fernandez A, Gershburg E. Rational Design of Live-Attenuated Vaccines against Herpes Simplex Viruses. Viruses. 2021;13(8):1637. Published 2021 Aug 18. doi:10.3390/v13081637
- 40.Stanfield BA, Bravo FJ, Dixon DA, Chouljenko VN, Kousoulas KG, Bernstein DI. Cross protective efficacy of the Non-Neurotropic live attenuated herpes simplex virus type 1 vaccine VC-2 is enhanced by intradermal vaccination and deletion of glycoprotein G. Vaccine. 2022;40(42):6093-6099. doi:10.1016/j.vaccine.2022.09.015
- 41. Dudek T, Knipe DM. Replication-defective viruses as vaccines and vaccine vectors. Virology. 2006;344(1):230-239. doi:10.1016/j.virol.2005.09.020
- 42.Nguyen LH, Knipe DM, Finberg RW. Replication-defective mutants of herpes simplex virus (HSV) induce cellular immunity and protect against lethal HSV infection. J Virol. 1992;66(12):7067-7072. doi:10.1128/JVI.66.12.7067-7072.1992
- 43.Krishnan R, Stuart PM. Developments in Vaccination for Herpes Simplex Virus. Front Microbiol. 2021;12:798927. Published 2021 Dec 7. doi:10.3389/fmicb.2021.798927
- 44. Wang Y, Zhang Z, Luo J, Han X, Wei Y, Wei X. mRNA vaccine: a potential therapeutic strategy. Mol Cancer. 2021;20(1):33. Published 2021 Feb 16. doi:10.1186/s12943-021-01311-z
- 45.Liu J, Fu M, Wang M, Wan D, Wei Y, Wei X. Cancer vaccines as promising immuno-therapeutics: platforms and current progress. J Hematol Oncol. 2022;15(1):28. Published 2022 Mar 18. doi:10.1186/s13045-022-01247-x